Cyclic Dimerization of 1,2-Unsaturated Maltotriose Derivatives with Iodinium Addition; One-pot Preparation of a Fully Methylated 2^A,2^D-Dideoxy-2^A,2^D-diiodocyclohexasaccharide

Nobuo Sakairi and Hiroyoshi Kuzuhara*

The Institute of Physical and Chemical Research (RIKEN), Wako-shi, Saitama 351-01, Japan

lodonium ion treatment of 1,2-unsaturated octa-O-methylmaltotriose having a sole hydroxy group at the 4"-position results in dimerization of the trisaccharide derivative with simultaneous cyclization, giving a fully methylated cyclohexasaccharide consisting of four α -D-glucopyranosyl residues and two 2-deoxy-2-iodo- α -D-mannopyranosyl residues.

Regio- and/or stereo-selective modification of cyclodextrins (CDs) have attracted much attention for both academic and industrial applications.¹ The characteristic structural feature of CDs, however, has narrowed the possibilities of such modification.

Recently, we succeeded in the cycloglycosidation of 1,2unsaturated maltohexaose derivatives with iodonium addition, giving mono(2-deoxy-2-iodo)cyclohexasaccharides.² Here we describe the one-pot synthesis of a 2^A ,2^D-dideoxy- 2^A ,2^D-diiodocyclohexasaccharide derivative starting from a trisaccharide glycal derivative by the extended application of the previous methodology. Thus, iodonium ion treatment of the trisaccharide starting material brought about simultaneous dimerization and cyclization to give a cyclic diiodo compound with two-fold symmetry.

The key trisaccharide glycal **5** was prepared from the thioglycoside $1^{\dagger} \{ [\alpha]_D^{24} + 58 (c \ 0.24, CHCl_3) \}$ derived from the known undeca-O-acetyl- β -maltotriose³ by the Lewis acid catalysed thioglycosideration as shown in Scheme 1. Thus, **1**

 $[\]dagger$ All new compounds gave satisfactory spectral data and elemental analyses.



Scheme 1 Reagents and conditions: i, NaOMe-MeOH; PhCH(OMe)2-TsOH, N,N-dimethylformamide (DMF), 60 °C, 20 mmHg, 6 h; $(MeO)_2SO_2-NaH$, DMF, room temp., overall 74%; ii, LiAlH₄-AlCl₃, Et₂O-CH₂Cl₂, room temp., 3 h, 93%; iii, $(MeO)_2SO_2-NaH$, DMF-THF, room temp., 88%; iv, lithium naphthalenide (8 equiv.), THF, $-80^{\circ}C \rightarrow$ room temp., overnight, 96%; v, IDCP (5 equiv.) molecular sieves 4 A, CH₂Cl₂, room temp., 1 day, 33%

was subjected to de-O-acetylation followed by O-benzylidenation and O-methylation, giving the 4,6-O-benzylidene derivative 2 { $[\alpha]_D^{28}$ + 74 (c 0.24, CHCl₃)}. Reductive ring opening of the benzylidene acetal under the conditions of Lipták et al.4 afforded the 6"-hydroxy derivative 3 {[α]_D²⁸ + 71 (c 0.22, CHCl₃), which was readily methylated to give the 4''-Obenzyl derivative 4 {(α]_D²⁸ + 88 (c 0.49, CHCl₃)} in 82% overall yield. Upon treatment with an excess of lithium naphthalenide⁵ under slighlty vigorous condition (-80 °C to room temp. overnight, in tetrahydrofuran (THF) under an argon atmosphere), 4 underwent a radical reductive elimination at the C-1 and C-2 positions with concomitant de-Obenzylation at C-4" to give the desired hydroxy glycal 5‡ $\{[\alpha]_D^{28} + 143 (c \ 0.26, CHCl_3)\}$ in almost quantitative yield.

Compound 5 was treated with iodonium di (sym-collidine) perchlorate⁶ (IDCP) in the presence of 4 A molecular sieves in CH₂Cl₂ at room temp. for 1 day. TLC of the reaction mixture showed that more than 3 compounds were produced, one of which moved faster on TLC than the starting material. The fast-moving products $\{[\alpha]_D^{28} + 101 \ (c \ 0.27, \ CHCl_3)\}$ was isolated as an amorphous powder in moderate yield by extractive work-up followed by column chromatography on silica gel (benzene-acetone, 3:1 v/v). Fast atom bombardment mass spectrometry (FABMS) of the product showed m/z1439.3 $[M + Na]^+$ and 1289.3 $[M - I]^+$ signals and determined that the product was a dimer of the trisaccharide. ¹H and ¹³C NMR spectroscopy revealed a simple pattern compatible with regular trisaccharide repeating units, which consisted of two α -D-glucopyranosyl residues [$\delta_{\rm H}$ 5.05 (d, J 3.4 Hz, 1-H), 5.03 (d, J 3.3 Hz, 1-H) and $\delta_{\rm C}$ 99.8, 99.7 (C-1)] and a 2-deoxy-2iodo- α -D-mannopyranosyl residue [δ_{H} 5.21 (s, 1-H-1) and δ_{C} 104.6 (C-1)]. Therefore, the product was elucidated as the 2^{A} , 2^{D} -dideoxy- 2^{A} , 2^{D} -diiodocyclohexasaccharide 6; yield of cyclization was 33%.

The cyclic hexasaccharide 6 possessed the ability to form a host-guest complex. Thus, the UV-VIS absorption spectrum of methyl orange in aqueous solution $(3 \times 10^{-5} \text{ mol dm}^{-3})$ at pH 1.0 showed a peak at 505 nm with a molar extinction coefficient of $\varepsilon 3.3 \times 10^4$ dm³ mol⁻¹ cm⁻¹, which decreased to $\epsilon 2.4 \times 10^4 \text{ dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ by addition of 6 (1 × 10⁻³ mol dm⁻³). The observed strong hypochromic effect suggested that 6 accommodated the methyl orange molecule in its cavity.7

By selective modification of the acyclic maltotriose starting material,3,8 the methodology presented here allows the efficient preparation of finely designed cyclohexasaccharides, which might be useful for construction of novel enzyme mimics or receptor models.

Received, 10th August 1993; Com. 3/04839J

References

- 1 W. Saenger, Angew. Chem., Int. Ed. Engl., 1980, 19, 344; I. Tabushi, Acc. Chem. Res., 1982, 15, 66; A. P. Croft and R. A. Bartsch, Tetrahedron, 1983, 39, 1417; S. Li and W. C. Purdy, Chem. Rev., 1992, 92, 1457.
- 2 N. Sakairi and H. Kuzuhara, J. Chem. Soc., Chem. Commun., 1992, 510; Chem. Lett., 1993, 1093.
- 3 A. Thompson and M. L. Wolfrom, J. Am. Chem. Soc., 1952, 74, 3612; N. Sakairi, M. Hayashida and H. Kuzuhara, Carbohydr. Res., 1989, 185, 91.
- 4 P. Fügedi, A. Lipták, P. Nánási and A. Neszmélyi, Carbohydr. Res., 1980, 80, 233; A. Lipták, J. Imre, J. Harangi, P. Nánási, and A. Neszmélyi, Tetrahedron, 1982, 38, 3721.
- 5 A. Fernandez-Mayoralas, A. Marra, M. Trumtel, A. Veyrières, and P. Sinaÿ, Carbohydr. Res., 1989, 188, 81.
- 6 R. U. Lemieux and A. R. Morgan, Can. J. Chem., 1965, 43, 2190.
- J. Szejtli, in Cyclodextrins and their Inclusion Complexes, Akademiai Kiado, Budapest, 1982, p. 162–175. 8 K. Takeo, T. Matsunami and T. Kuge, *Carbohydr. Res.*, 1976, **51**,
- 73; N. Sakairi and H. Kuzuhara, Carbohydr. Res., 1993, 246, 61.

[‡] Selected 1H NMR (400 MHz, CDCl₃) data: δ 2.86 (s, 1 H, OH), 3.22 (dd, 1 H, J, 3.5, 9.8 Hz, 2-H), 3.29 (dd, 1 H, J 3.4, 9.8 Hz, 2-H), 3.32, 3.35, 3.39, 3.40, 3.49, 3.52, 3.58, 3.63 (8 × s, 8 × 3H, 8 × Me), 4.83 (dd, 1 H, J 2.5, 6.3 Hz, 2¹-H), 5.59 (d, 1 H, J 3.9 Hz, 1-H), 5.66 (d, 1 H, J 3.9 Hz, 1-H-1), and 6.44 (d, 1 H, J 5.9 Hz, 11-H).

[§] Selected spectral data: ¹H NMR (400 MHz, CDCl₃); & 3.15 (dd, 2 H, J 3.4, 9.8 Hz, 2-H), 3.18 (dd, 2 H, J 3.1, 9.5 Hz, 2-H), 3.38, 3.39, 3.41, 3.46, 3.47, 3.62, 3.63 (s, Me), 3.84 (dd, 2 H, J 4.0, 10.7 Hz, 6-H), 3.91 (dd, 2 H, J 4.3, 10.3 Hz, 6-H), 4.57 (br d, 2 H, J 4.3 Hz, 2¹, 2⁴-H), 5.03 (d, 2H, J 3.4 Hz, 1-H), 5.05 (d, 2H, J 3.4 Hz, 1-H), and 5.21 (br s, 2H, 11, 14-H); 13C NMR (67.8 MHz, CDCl₃); 8 33.2, 57.6, 57.9, 58.0, 59.0, 59.1, 59.2, 61.0, 61.7, 70.7, 70.9, 71.1, 72.0, 73.2, 77.2, 80.3, 80.8, 81.0, 81.2, 82.0, 82.1, 82.5, 99.7, 99.8 and 104.6.